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Prevalence of Salmonella in Human and Foods of Animal Origin with Antibiotic Resistance Patterns of Isolated Bacteria in Dhamar Governorate, Yemen

Ahmed M. Al-Khadher and Samiha N. Maglas

Veterinary Medicine department., Faculty of Agriculture & Veterinary Medicine, Thamar University, Dhamar, Yemen.

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-ABSTRACT

Salmonellosis is a food borne zoonotic disease and one of the major the public health problems. Assessment of Salmonella contamination risk In Food of Animal Origin at Dhamar governorate is the main objective for this study. A total of 188 samples of animal food products, including, white meat, red meat, eggs and milk were collected from slaughterhouses, local markets and cows. In addition, a total of 35 stool and 45 blood samples from human were collected. All Samples were collected in aseptic manner, brought to laboratory and subjected to bacteriological, biochemical and serological tests accordingly. assessment For sensitivity of isolated

species to antibiotics, ten antibiotic drugs were used for this purpose. The results revealed that, out of 188 samples examined and tested for salmonella species, 37(20%) were positive. The positive rates according to sample-type were 0.11%, 0.14%, 0.05%, 0.11%, 0.05%, 0.16%, 0.19%, 0.14%, and 0.05% for native beef, imported beef, camel meat, minced meats, poultry meat, fish meat, table eggs, and raw milk respectively. In human, out of 35 stool and 45 blood samples examined, 8(22.8 %) and 22(48.9%) were positive for salmonellosis respectively. The serovars identified were; typhi (O) 49% typhi (H) 36% Paratyphi (B) 24%, but Paratyphi (A) hasn't found. All isolated chloramphenicol salmonella sensitive ciprofloxacin, nalidixic acid were to gentamicin, and intermediate resistant for Erythromycin and Tetracycline, but complete resistant to Colistin, Doxycycline, Penicillin and Lincomycin antibiotics. This study confirmed that, food animal origin at Dhamar governorate are highly contaminated with some Salmonella spp and those species representing the main source of salmonella infections in patients in Dhamar governorate. Further studies on Salmonella

spp. Isolation, spread tracking through production cycle of farm animals, and control measures are recommended.

Key words: Animals, Human, Dhamar, Prevalence, Sallmonella.

INTRODUCTION

Salmonellosis is a food borne zoonotic disease of primary concern in developed, as well as developing countries. spread of this disease is favored by a variety of animal reservoirs and a wide commercial distribution of both animals and food products. This disease is among one of the major public health problems terms of socio-economic impact (Mushtag et al., 2008). About 95% of affected human with salmonella were infected by consumption the contaminated animal food origin such as meat, poultry, eggs, milk, seafood and fresh produce (Callaway et al, 2000). They cause a wide range of clinical illness: enteric fever, gastroenteritis, and bacteraemia, particularly in infants and in immunocompromised patients (Fluit, **2005**).Several studies from different countries have been reported prevalence and the relationship between Salmonella spp., foods of animal origin and public health problems (Rasrinual et al., 1988; Boriraj et al., 1997; Boonmar et al., 1998); Whereas, in Yemen, there are few studies about Salmonellosis that were mainly focusing on human infection (Al-Haddad, 2004 and Saleh, 2010) and local cooked foods (AL-Ammari, 2012). Reports revealed that, in 2009, about 39,770 of Typhoid and Paratyphoid cases were recorded in Yemen (CSO, 2009).

In Dhamar governorate, Salmonellasis is common and has been diagnosed as enteric fever in patients (16.4%) and food poising cases (15.2%) (Taha et al., 2013). However, no data available in compiled form reporting the current status of salmonellosis and role of animal food origin as source for human infections. Therefore, this study was carried out determine prevalenceof to salemonella species and assess their susceptibility and resistance to some antimicrobial drugs.

MATERIAL AND METHODS:

The study had been conducted in Dhamar governorate through April until December 2013. A total of one hundred and eighty eight(188) samples of different animal product food were collected from different regions of Dhamar governorate includeds laughters, local markets and cows. also 35 clinical specimens (human stool samples) and 45 of blood samples had taken from patients with suspected enteric fever whom were attending General Dhamar hospital to determine the incidence of Salmonella infection(Table 1).

Table 1:Samples from food of animal origin

Type of raw samples	Red meat				White meat			eggs	milk	Human samples		
species	native beef	imported beef	Camel meat	Minced meats	Freeze chickens	fresh chickens	Fish meat	table eggs	Raw Milk	stool	serum	Total
No. of collected samples	17	16	5	30	10	20	30	30	30	35	45	268

For meat samples, Approximately 25g of meat pieces were harvested from different parts of carcasses and each sample was aseptically placed into a sterile vacuum bag.Milk samples was collected by pouring approximately 50 ml into a sterile specimen cup. Blood samples were inoculated in sterile test tube for sera collection while stool samples were placed in selenite and tetrathionate broth.

Collected samples were taken, properly labeled and brought to, Faculty of Agriculture and Veterinary Medicine laboratories on the day of collection in Chilled ambient using thermos supplied with ice.

For bacteriological analysis, samples preparing, grinding and homogenizing of meats were carried out in sterile conditions, and then, 25g of meat or stool, 10 ml of milk and 10 ml of egg yolk samples were added to 100 ml of selenite enrichment broth (**Himedia**®) and incubated at 37°C for 24h under aerobic conditions.

The positive growth with selenite broth were transferred to selective media salmonella shigella agar (SSA) (**Himedia**®) and then streaked on Mac- Conkey's agar (**oxoid**®). After 24 h incubation at 37°C, two or more ofsuspect colonies from each agar plate (non-

lactose fermented coloniesin Mac- Conkey's agar), that revealed typical physiognomies of Salmonella were pickedout and submitted to biochemical analysis byusing sulfite-Indole-Motility agar (SIM) (Himedia®),Oxidase, Catalase, Triple Sugar Iron Agar (TSI) (Himedia®), Ureas (Himedia®) and simmon's citrate tests according to (WHO, 2010)and gram staining was carried outfor all apparent Salmonella colonies to morphological identification according to (Benson, 2001). Human serum samples were submitted to Slid

(Widal) test (**Himedia**®) against s. typhi 'O', and 'H', and s. paratyphi A 'H' and s. paratyphi B 'H' antigens to determine the serovars present in human serum samples.

Sensitivity tests were conducted for confirmed *Salmonella* isolates using Muller-Hinton agar plates (**Himedia**®) by disk diffusion pattern using 10 types of antibiotic disks (**Himedia**®)as shown in table 2.

Table 2: antibiotic discs and their potency used in Sensitivity tests of isolated salmonella

Antibiotic	Lincomycin (L)	Penicillin (P)	Tetracycline (T)	Nalidixic Acid (NA)	Gentimycin (G)	Erythromycin (E)	Doxycycline (D)	Colistin (CL)	Ciprofloxacin (CIP)	Chloramphenicol (C)
Disk potency	2 μg	10 μg	30 µg	30 µg	10 μg	15 μg	30 µg	10 μg	5 μg	30 μg

RESULTS:

The results showed that, positive samples of food origin samples with *Salmonella* were 37 (20%) out of (188) different samples, individual percentage for each kind ranged from 0.05 % of raw milk to 0.19% of fish meat samples (**figure, 1**). Whereas, 8 of total 35 collected samples of human stool (22.8 %),were positive

salmonella, and human blood samples had 48.9% positive serological results (**Table 3**).

About half of collected blood samples (22) showed positive reaction with Widal test against. typhi 'O', and 'H', and s. paratyphi A 'H' and s. paratyphi B 'H' antigens, and theserovars identified were; typhi (O) 49% typhi (H) 36% Paratyphi (B) 24%, but Paratyphi (A) hasn't found(**figure**, **2**).

The microscopic future of gram stained isolates, exhibited a typical morphological characteristics of salmonella spp. in single, coupled, or clumped, gram positive road cells (**Figure 3**).

As showmen in **Table (4)**, Isolated Salmonella were highly resistant tocolistin,

doxycycline, penicillin, and lincomycin but moderate resistant was found against tetracycline and erythromycin, however, All isolated Salmonella were susceptible to ciprofloxacin, gentamycin, chloramphenicol and nalidixic acid.

Table 3:Salmonella positive of Human samples which collected at General Dhamar Hospital.

Human specimens	Number of samples		Po	Positive%			
stool	35			22.8			
blood	45 22		Paratyphi (A)	Paratyphi (B)	(O) 22	typhi (H)	48.9%
total	80	30					37.5

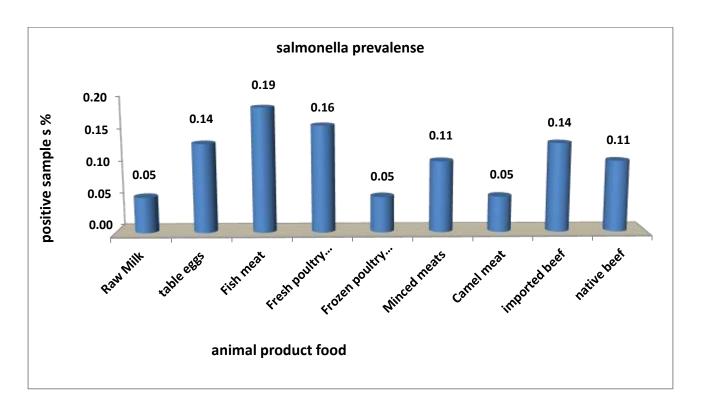


Figure 1: The distribution rates of positive Salmonella samples.

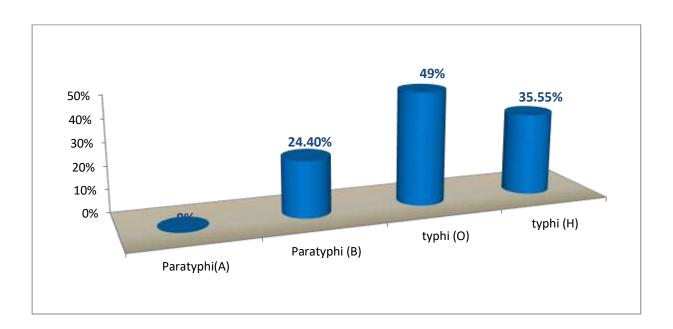


Figure 2: The positive widal test percentage of each kind of samples.



Figure 3: Salmonella gram stain (Gram negative, short rod).

Table 4: Zones of clearing for different antibiotics against isolated salmonella

Antibiotic	Disk potency	Sensitivity				
		Inhibition zone	Result			
		diameter / mm				
Chloramphenicol (C)	30 μg	27 - 30	Susceptible			
Ciprofloxacin (CIP)	5 μg	25	Susceptible			
Colistin (CL)	10 μg	-	Resistant			
Doxycycline (D)	30 μg	-	Resistant			
Erythromycin (E)	15 µg	14 – 11	Intermediate			
Gentimycin (G)	10 μg	17 - 20	Susceptible			
Nalidixic Acid (NA)	30 μg	18 - 20	Susceptible			
Tetracycline (T)	30 μg	16	Intermediate			
Penicillin (P)	10 μg	-	Resistant			
Lincomycin (L)	2μg	-	Resistant			

DISCUSSION:

In Yemen, high contamination of cocked and ready-to eat food with salmonella, also, the incidence of Salmonella infection in peoples suffering from enteric fever and food poisoning had detected (AL-Ammari, 2012 and Taha et al., 2013), but no satisfied investigation or data about the mainsources or reservoirs of these infections. This study designed for solution of this riddle. The present results showed that the highest contamination rates were in fish meats(19%), followed by fresh poultry meats (16%), which agree with many former studies (Dhaher, et al, 2011). The cause of this phenomenon may due to high contamination of poultry feed

(Jones and Richardson, 2004, and EFSA, 2008a.)in addition to contamination through transportation and processing in unplanned and muddled local slaughters that poorly equipped don't followed the minimal hygiene precautions.whereas, the mentioned two types of foods are public and the cheapest source of protein in Yemen, hence, in many developed countries, the chicken meats are the main source of human infections, they make a potential threaton nation public health .(Al-Matar et al, 2005 and Xia et al., 2009). The table eggs were at the third rank of positive samples which were 0.14% this rate is constant with Herdberg, (1993) ICMSF.

(1996)., and Poppe, et al., (1998), whose found positive samples ranged between (%0.4) and (0.17 %). The lowest rates of positive samples were in raw milk (0.05%), which agreed with AL-Ammari, (2012) who found similar results in animal product cocked food. Also camel meats and frozen chicken samples had low rates (0.05) of salmonella positive, this ratified with Lake et al., (2002), who found low contamination of child chicken but for camel meats, in present study, this may due to the paucity of this type of meat in Dhamar that lead to low number of collected samples.

The high percentages of positive stool samples exposed a high incidence of salmonellosis (22.8) in peoples whom were attending General Dhamar hospital. These findings were similar to those reported by Al-Haddad, (2004) and saleh, (2010). These results may be due to the most of studied patients had a history of direct contacts with domestic animals or its products that may be the main source of their infections(Al-Matar et al., 2005 and Xia et al., 2009).

The positive serological results of human blood, revealed that typhi (O) was the predominant serovars (49%) followed by typhi (H) (36%) whereas, Paratyphi (B) was the lowest (24%), and no positive results had found with Paratyphi (A), these results had Approved With **Taha et al.**, (2013). The high prevalence

of Salmonella Typhi may be due to the fact that Salmonella Typhi is spread predominantly within the household, whereas Salmonella Paratyphi is mainly transmitted outside the home(Rice et al., 2003 and Massi et al., 2005).

Isolated Salmonella were highly resistant tocolistin, doxycycline, penicillin, and lincomycin. This outcome is consistent with Taha et al., (2013) and Gautam (2002) this resistance may be due to miss-used of antibiotics without those medical superintendence in Yemeni's animal and poultry farms as treatment or growth Salmonella were promoters.All isolated susceptible to ciprofloxacin, gentamycin, chloramphenicol and nalidixic acid.

Conclusions and recommendations:

Salmonella was present in allsample kinds that were examined in Dhamar governorate. This study has confirmed that, animal food origin at Dhamar province, were highly contaminated with Salmonella and believed that, those products (particularly, chicken and fish meat, which are public and the cheapest source of protein in Yemen) are the main source of salmonella infection in Dhamar's people.

From our results, it recommended that, more studies on spread tracking of Salmonella spp. and control measures in the production cycle of farm animals are required. The good hygiene, good cocking for animal product foods and medical superintendence on animal farms and slaughters should be directed to controlling the spread of Salmonella, and reduce the number of Salmonella infections.

REFERENCES

Al-Haddad, A. M. (2004): Serodiagnosis of Infectious Diseases among Patients admitted to IbnSina Central Teaching Hospital in Al-Mukalla City-Yemen. *Journal of Hadhramout University; 3 (6): 173-179.*

AL-Mashhadany, D. A., 2008. The Hygienic importance of Salmonella in Red meat, Journal of Thamaruni., I, (1):17-25.

Al-MatarAimanHamed, Khalil Al-Shawabkeh and Hana Zakaria (2005)., Prevalence of Salmonella in Broiler Chicken Carcasses in Jordan. Studies onf agriculture sciences. Vol. 32, no. 2, pp:267-277.

Benson (2001): Microbiological Applications Laboratory Manual in general Microbiology. Part four. Culture Methods. 23. Bacterial Population Counts. 8thedition. (Ed): Fornango J. and Smith J. The McGraw–Hill Companies.

Boonmar, S., Bangtrakulnonth, A., Pornruangwong, S., Samosornsuk, S., Kaneko, K., Ogawa, M., 1998. Significant increase in antibiotic resistance of Salmonella isolates from human beings and chicken meat in Different countries. Veterinary Microbiology 62, 73–80.

Boriraj, V., Bangtrakulnonth, Pornruangwong, S., Saitanu, K., 1997. Demographic on Salmonella data enteritidis infection in Different 1990-1995. countries. Southeast Asian Journal of Tropical Medicine and Public Health 28, 774–780.

Organization, Central **Statistical** Yearbook. (CSO). (2009): Statistical Republic of Yemen. Ministry of planning &int.Cooperation. Central statistical organization. General management of dissemination publication. & 39th edition.

Dhaher F. H. D. H. Awni M N. R. Mahmood M.M. Jamil and H. S. Rasheed (2011) Isolation and Diagnosis of Salmonella in Animal Origin Food, Import feed in Baghdad Local Markets and Local Poultry Farms. Iraqi journal for market researches and consumptive prevention, vol. 3; no.5.,pp.1-19.

European Food Safety Authority, (EFSA). (2008a): Microbiological risk assessment in feeding stuffs for food-producing animals. Scientific Opinion of the Panel on Biological Hazards. The European Food Safety Authority Journal; (EFSA-Q-2007-045). 720: 1-84.

European **Food** Safety Authority, (EFSA). (2008b): Quantitative Microbiological Risk Assessment Salmonella in Meat1: Source Attribution for Human Salmonellosis from Meat. The European Food Safety Authority Journal: 625: 1-32.

Fluit CA (2005).Towords more virulent and antibiotic-resistant Salmonella?. FEMS Imm. Med. Microb., 43: 1-11.

Gautam V (2002). Sensitivity pattern of Salmonella serotypes in Northern India. Brazilian Journal of Infectious Diseases, , 6:281–287.

Herdberg, C.W., David, M.J., White, K.E., McDonald, K.L. and Osterholm, M.T. 1993. Role of Egg Consumption in Sporadic Salmonella enteritidis and salmonella typhimurium Infections in Minnesota. Journal of Infectious Diseases, 167: 107-11.

International Commission on Microbiological Specification of Food. ICMSF. (1996). Egg and Egg Products. In Microorganism in Food 6: Microbiology of Food Commodities. London: Chapman and Hall. 475-520.

Jones, F. T. and Richardson, K. E. (2004). Salmonella in commercially manufactured feeds. Poultry Science. 83: 384-391.

Lake, R.; Hudson, A. and Cressey, P. (2002). Risk Profile: Salmonella (Non-Typhoid) in Poultry (Whole and Pieces). Acrown Research Institute, Client report FWO 212. ESR, New Zealand.

Massi, M. N; Shirakawa, T; Gotoh, A; Hatta, M. and Kawabata, M. (2005). Identification Sequencing and of EntericaSerotype Salmonella **Typhi** Obtained from **Patients** Isolates Perforation and Non- PerforationTyphoid Fever. International Center for Medical Research. Kobe University School Medicine. The Southeast Asian Journal of Tropical Medicine and Public Health: 36(1): 118- 122.

Mushtaq, H., MI Hussain, B Shahzadi, M Shaheen, MS Mahmood, A Rafique and M Mahmood-ul- Hassan, 2008. Occurrence of

some zoonotic microorganisms in faecal matter of house rat (Rattusrattus) and house mouse (Musmusculus) trapped from various structures. Pakistan Vet J, 28(4): 171-174.

Poppe, C.; Duncan, C. L. and Mazzocco, A. (1998). Salmonella contamination of hatching and table eggs: a comparison. Can. J. Vet. Res. 62: 191-198.

Quirke, A. M., N. Leonard, G. Kelly, J. Egan, P. B. Lynch, T. Rowe, and P. J. Quinn. 2001. Prevalence of Salmonella serotypes on pig carcasses from high- and low-risk herds slaughtered in three abattoirs. Berl. Muench. Tieraerztl. Wochenschr. 114:360-362.

Rasrinual, L., Suthienkul, O., Echeverria, P., Taylor, D., Seriwatana, J., Bangtrakulnonth, A., Lexomboon, U., 1988. Foods as source of enteropathogens causing childhood diarrhea in Different countries. American Journal of Tropical Medicine and Hygiene 39, 97–102.

Rice DH et al (2003).Household contamination with Salmonella enterica. *Emerging Infectious Diseases*, , 9:120–122.

Saleh, M. G. S. (2010):Investigation on Salmonella spp. in patients suffering from fever and food poisoning in Thamar city-Yemen. (MCs), Sana'a University, Faculty of Science. Yemen.

Taha R.R.,S.M. Alghalibi and M.G. SaeedSaleh; 2013 Salmonella spp. in patients suffering from enteric fever and food poisoning in Thamar city, Yemen. *EMHJ* • *Vol.* 19 No. 1 •

World Health Organization, (WHO). (2010): Laboratory Protocol Isolation of Salmonella spp. 5th Ed. From Food and Animal Faeces. WHO Global Foodborne

Infections Network (formerly WHO Global Salm-Surv): 1-18.

Xia, X; Zhao, S; Smith, A; McEvoy, J; Meng, J. and Bhagwat, A.A. (2009): Characterization of Salmonella isolates from retail foods based on serotyping, pulse field gel electrophoresis, antibiotic resistance and other phenotypic properties. International Journal of Food

Microbiology;129:93-98.www.elsevier.com/locate/ijfoodmicro.

AL-Ammari Y. N. M (2003). Incidence and Distribution of Salmonella Serotypes in Some Local Foods in Sana'a. MSc. Thesis, Submitted to the Department of Biology, Faculty of Science, Sana'a University.

انتشار السالمونيلا في الإنسان والأغذية ذات المنشأ الحيواني ودراسة حساسيتها للمضادات الحيوية في محافظة ذمار / اليمن

أحمد محمد الخضر و سميحة ناصر مقلس

قسم الطب البيطري، كلية الزراعة والطب البيطري، جامعة ذمار، ذمار، اليمن.

الخلاصة:

تعتبر السالمونيللا من الامراض الغذائية المشتركة وتشكل أحد المشاكل الرئيسية للصحة العامة. أجربت هذه الدراسة بهدف تقييم خطر انتشار السالمونيلا في الأطعمة ذات المنشأ الحيواني في محافظة ذمار. جمعت عدد مائة وثمان وثمانون عينة (١٨٨) من أغذية المنتجات الحيوانية المختلفة حيث جمعت من المسالخ والمجازر المحلية والبقالات وجمع الحليب من الأبقار. كذلك تم جمع خمس وثلاثون (٣٥) عينة من البراز البشري. جرى تجهيز هذه العينات وتحضيرها وخضعت للفحص البكتيري وتم تأكيدها بالاختبارات البيوكيميائية لتحري وجود السالمونيللا. سحبت خمس وأربعـون عينــة دم مــن الحــالات البشــربـة المشــتبهه لتحديــد مــا اذا كانــت مصــابـة بالســالمونيلا باســتخدام الاختبــار السريع على الشريحة (اختبار وايدل). خضعت عشرة أنواع من المضادات الحيوبولأختبار حساسية عزلات السالمونيلا المدروسة لها. أضهرت النتائج وجود السالمونيللا في في حوالي ٢٠ % من مجموع العينات المدروسة، توزعت النتائج الموجبة على الأغذية المختلفة بحيث كانت: ١١٠، %، ١٤٠%، ٥٠,٠٥%، ٢٠,١١، %، ٥٠,٠٠%، ٠,١٩ %، ١٤٠%، و ٠,٠٠ % للحم البقرالمحلى، لحم البقرالمستورد، لحم جَملِ، اللحم المفروم، لحوم الدجاج، لحوم السمكِ، بيض المائدةِ، والحليب الخام على التوالي. ٢٢,٨ % مِنْ مجموع عينات البراز البشري، كانتْ إيجابية تحتوي على السالمونيلا ، بينماأظهرت عينات الـدمّ البشـري نتـائج ايجابيـة فـي٤٨٫٩ % منهـا بالاختبـارات المصـلية، شـكّل الـنمط المصلى (O) للسالمونيلا تايفي ٤٨% والنمط المصلى (H) ٣٦% بينما وجد النمط المصلى للسالمونيلا نظيرة التايفويد ٢٤% ولم يوجد النمط المصلى (A) للسالمونيلا نظير التايفويد. أضهرت السالمونيلات المعزولة حساسية للكلورامفينيك ول والسيبر وفلوكساسين والناليديكسيك أسيد والجنتامايسين، وحساسية متوسطة للإربثر ومايسين والتتراسيكلين، ولكنها اظهرت مقاومة كاملة للكولستين والدوكسيسيكلين والبنسلين واللينكومايسين. لقد أكدت هذه الدراسـة أن الأطعمـة ذات المنشـأ الحيـواني فـي محافظـة ذمـار تحتـوي علـي تلـوث عـالي بالسـالمونيلا وبعتقـد أن هـذه الأطعمـة هـي المصـدر الرئيسـي لعـدوي السـالمونيلا لـدي سكان ذمـار . وبوصـي بمزبـد مـن الدراسـات لتتبـع انتشـار السالمونيلا وطرق السيطرة عليها في في حلقات الانتاج للحيوانات الزراعية.